

Insect Assemblage as Bioindicators for Stream Quality within Two Sacred Groves in Ghana

Victor Parry¹, Roger Sigismund Anderson², Samuel Adu-Acheampong^{2,3*},
Mukundi Mukundamago^{4,5}, Emmanuel Robert Blankson^{3*}, Rosina Kyerematen^{2,3}

¹Future Water Institute, University of Cape Town, Cape Town, South Africa

²African Regional Postgraduate Programme in Insect Science, University of Ghana, Accra, Ghana

³Department of Animal Biology and Conservation Science, University of Ghana, Accra, Ghana

⁴International Centre of Insect Physiology and Ecology, Nairobi, Kenya

⁵Department of Earth Sciences, University of the Western Cape, Bellville, South Africa

Email: *samueladuacheampong81@gmail.com, *erblankson@gmail.com

How to cite this paper: Parry, V., Anderson, R. S., Adu-Acheampong, S., Mukundamago, M., Blankson, E. R., & Kyerematen, R. (2026). Insect Assemblage as Bioindicators for Stream Quality within Two Sacred Groves in Ghana. *Journal of Geoscience and Environment Protection*, 14, 64-88.

<https://doi.org/10.4236/gep.2026.145006>

Received: April 23, 2026

Accepted: May 23, 2026

Published: May 26, 2026

Copyright © 2026 by author(s) and Scientific Research Publishing Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

Sacred groves are patches of relict forests set aside by communities for traditional, religious and conservation purposes. However, freshwater ecosystems in these sacred groves are impacted by anthropogenic activities, due to lack of enforcement of traditional edicts to check encroachment. This study measured the effect of stream dynamics (stream size and water quality) on aquatic insect diversity and by extension to assess the stream health in two sacred groves-Abiriw and Odumante-in Ghana. Insects were sampled across rainy and dry seasons using multiple standard collection methods and compared between these sacred groves via sampling different microhabitats associated with the different human impacts using Dip net and Surber Stream-bottom sampler to ensure a representative sampling at all places within the streams. We also used Eureka Sub3 multiparameter water-quality multiprobe to measure the physico-chemical parameters of the streams. Results: The study recorded 2767 aquatic insects classified into 65 families and 11 orders. The results showed that abundance did not differ significantly between groves, but family richness was higher at Odumante (60 families) compared to Abiriw (46 families). The results further revealed that seasonality influenced insect diversity with both abundance and richness significantly higher in the dry season in both sacred groves. There was low representation of Ephemeroptera, Plecoptera, and Trichoptera (EPT) and high Hemiptera dominance suggesting moderate to high environmental stress especially in these streams. Conclusions: Based on the results we can infer that streams in the two sacred groves have been impacted by anthropogenic activities with a resultant change in stream dynamics and a profound influence on aquatic insect communities, especially at Odumante with implications for in-

tegrated conservation of sacred groves freshwater ecosystems. We recommend that conservation efforts must address watershed-level waste management and farming activities along the riparian zones in sacred groves to protect aquatic ecosystems.

Keywords

Abiruw, Biodiversity, Bioindicator, Odumante, Stream

1. Introduction

Sacred groves are remnant forest patches preserved through cultural or religious practices, which may serve as refugia for biodiversity in fragmented landscapes (Aniah & Yelfaanibe, 2018; Tatay & Merino, 2023; de Souza, 2025). In Ghana, these sacred groves protect culturally significant trees, vegetation, and water bodies dedicated to deities or ancestral spirits (Anokye, 2022) and as a result, local norms are used to enforce their protection via taboos, integrating spiritual and ecological values (Diawuo & Issifu, 2017; Kosoe et al., 2020). Historically, sacred groves have been used as biodiversity conservation hubs (Patole, 2022; Sarfo-Adu et al., 2022; Kujur et al., 2025; Ziblila et al., 2025), yet contemporary anthropogenic pressures such as encroachment, farming, and mining threaten their integrity (Nganso et al., 2012; Boamah, 2020; Oppong, 2023; Asante, 2024). It is estimated that there are about 2000 - 3200 sacred groves in Ghana with close to 80% occurring in the southern half of the country (Gordon, 1992; Osei et al., 2018; Batame et al., 2023). Generally, the main objective for establishing sacred groves in communities in Ghana is to protect forests surrounding environmentally sensitive areas. The size of sacred groves in Ghana ranges from hundreds of hectares of large forest (>100 ha) to small plots (<0.5 ha) containing single trees or few stones (Gordon, 1992; Franks et al., 2017). Because of their perceived attachment to some deities or ancestral spirits, sacred groves are referred to in different communities, such as nananompow (ancestral grove or royal Mausoleum) (Adarkwa-Dadzie, 1997), abosompow or asoneyeso (shrine), mpanyinpow (ancestral forest), and nsamanpow (burial grounds) by the Akans (Ntiamoa-Baidu, 1995; Attuaquayefio & Fobil, 2005).

Sacred groves serve ecological and socio-cultural functions like preserving virgin forests, serving as a refuge for rare and endemic local biodiversity. They also serve as sources of herbs for medicinal, social and religious purposes (Asokan et al., 2015; Doffana, 2017; Ray & Ray, 2020). Because of the linkage between traditional beliefs and deities or ancestral spirits, disobedience or disregard for traditional laws (taboos) regarding the encroachments of sacred groves attract severe punishments for culprits. Such punishments include offering animals as sacrifice and the performance of traditional rites to appease the local deity to avoid ill-health, and deaths, although this is often ineffective in modern Ghana unlike centuries ago (Kwarteng, 2015; Ikenyei & Lawal, 2019). Anthropogenic activities have

been reported to have been a major cause of biodiversity loss in the world although the bulk of research in this area within sacred groves is mainly focused on botanical, Ethno-botanical or socio-cultural functions (Kosoe et al., 2020; Sarfo-Adu et al., 2022; Kumar et al., 2022; Gyedu et al., 2025). Notwithstanding this general situation, there have been some studies on insect diversity assessment in sacred groves as reported by Nganso et al. (2012), Mukherjee et al. (2018), and Mondal & Mondal (2024).

The class Insecta contains some important groups with several characteristics that make them suitable for ecosystem health monitoring (Mensah et al., 2018; Nnoli et al., 2019; Menta & Remelli, 2020; Parikh et al., 2021; Chowdhury et al., 2023). They inhabit all habitat types and play major roles in the functioning and stability of both terrestrial and aquatic ecosystems (Adler & Footitt, 2009). For instance, it has been reported in several studies that diversity and abundance of grasshoppers (Adu-Acheampong et al., 2016; Adu-Acheampong & Samways, 2019; Khan et al., 2023; Mariottini et al., 2024), butterflies (Sharma & Sharma, 2017; Kyerematen et al., 2018a, 2018b; Ismail et al., 2020; Pallottini et al., 2023) beetles (Mukweho et al., 2017; Carvalho et al., 2020; Karpiński et al., 2021; Makwela et al., 2023), and ants (Carvalho et al., 2020; Zina et al., 2021; Lutinski et al., 2024) can be used as good indicators of ecosystem health. The most common features that make insect ideal for bioindication are their ubiquitous occurrence, high species richness, basic sedentary nature, sensitivity to environmental pollutants, easy and inexpensive sampling and well-described taxonomy for families and genera (Resh et al., 2004; Bonada et al., 2006; Anderson et al., 2025). In line with that, aquatic insects have been widely used for measuring health within freshwater ecosystems in the past few decades (Hodkinson & Jackson, 2005; Bonada et al., 2006; Nicacio & Juen, 2015; Mensah et al., 2018; Nnoli et al., 2019). Orders like Ephemeroptera, Plecoptera, and Trichoptera (EPT) are highly responsive to changes in water quality and as such have been used in bioassessment frameworks globally (Bonada et al., 2006; Miguel et al., 2017; El Yaagoubi et al., 2025; Indraswari Suhri et al., 2025).

Recent evidence however suggests that the structure of aquatic insect communities is not solely shaped by pollution or land use, but also by intrinsic ecological factors such as seasonality and stream dynamics, such as stream size and physico-chemical parameters (water quality) of the stream (Kasangaki et al., 2008; Ohler et al., 2023; Pedrosa Guimarães et al., 2025). For instance, a study conducted at the University of Ghana and such similar studies elsewhere exploited the sensitivity of Odonata to changes in the freshwater ecosystem in generating baseline information about the current state of aquatic systems (Acquah-Lampsey et al., 2013; Miguel et al., 2017). Odonata diversity associated with freshwater bodies has further been exploited as a tool for predicting the potential future changes in habitat quality (Acquah-Lampsey et al., 2013; Miguel et al., 2017; Manu et al., 2023; Datto-Liberato et al., 2024). While well-documented globally, seasonal multi-taxon assessments remain under-represented in West African sacred grove literature. Seasonal variation in tropical streams, particularly the shift between rainy

and dry seasons, is known to cause profound changes in aquatic insect communities. Studies from Ghana and other tropical countries (e.g. [Sánchez-Argüello et al., 2010](#); [Kyerematen & Gordon, 2012](#); [Agodzo et al., 2023](#)) consistently show peaks in abundance and diversity during the dry season, often due to concentration of individuals in residual pools and more stable water conditions. Similarly, stream dynamics affects habitat heterogeneity, with smaller streams often experiencing greater environmental variability and stronger community shifts ([Maloney et al., 2011](#); [Brasil et al., 2020](#); [Sankone et al., 2023](#)).

Despite studies done in sacred groves in Ghana, there is no study in these streams that seeks to measure the effect of stream dynamics (e.g. stream size and water quality) on aquatic insect diversity and by extension, assess the stream health in the two sacred groves, Abiriw and Odumante, in Ghana. And so, this study could serve as a baseline study to kick start other studies that will lead to comprehensive studies on stream health within these two sacred to be used to inform conservation strategies in the same. In line with that this study investigates how seasonality and stream dynamics affected by anthropogenic activities interact to shape aquatic insect diversity within two sacred groves in Ghana as a measure of stream health within these sacred groves.

We hypothesized that seasonality and stream dynamics including localized disturbances, have a profound influence on aquatic insect diversity within Abiriw and Odumante sacred groves. This study contributes to the growing body of literature advocating for integrated ecological and cultural conservation of sacred groves using aquatic insect diversity as a bioindicator in measuring the health within aquatic ecosystems. Specifically, the study determined aquatic insect diversity in the two sacred groves, assessed the seasonal variation in insect communities and used the information to evaluate stream health within these sacred groves.

2. Materials and Methods

2.1. Study Area

The Abiriw sacred grove (5°48'N, 0°06'W) is in Abiriw and covers an area of 400 m² in the Akwapim North District of the Eastern Region of Ghana. The area falls within the Moist Semi-Deciduous Forest vegetation zone which is characterized by two rainfall peaks annually. This study site has rich floral diversity with climbers being most dominant. Most of the trees within this area grow up to 37 m high ([Ayivie, 2005](#)). Some of the dominant tree species within this sacred grove are *Antaris toxicaria*, *Trichilia monadalpha*, *Bahia pubescens*, *Viten gradifolia* and *Macaranga bateri* ([Ayivie, 2005](#)). Even though there was no active human activity within this stream, there were both biodegradable and nonbiodegradable wastes found at the base of the waterfall of the stream as they are carried from surrounding villages along the upper reaches of the stream to the sacred grove as indication of human activity. The adjacent lands to the Abiriw stream were semi natural lands from which the villagers collect firewood. The stream flow in Abiriw sacred grove is a first-order or low-order (1 to 2) stream in the Strahler

stream order system.

The second stream for this study lies at the edge of the Odumante Sacred grove. The Odumante Sacred grove (5°28'N, 0°18'W) is located at Kitase-Aburi in the Akwapim South District of the Eastern Region of Ghana. This Sacred grove covers an area of 250 m². This study site also has the same environmental factors such as rainfall, temperature, rainy seasons and vegetation type as Abiriw. Both study sites experience bimodal rainfall (major rainy season occurs between April and June, while the minor one falls within September and October). The annual rainfall ranges from 1300 mm to 1800 mm. The stream flows through the village and passes close to the Sacred grove. There were some localized disturbances in this sacred grove like active firewood gathering and pollution of the stream by both biodegradable and non-biodegradable items coming from farming and other activities emanating from the inhabitants within the village who also depend on the stream as a source of water. There were cultivated oil palm and plantain trees adjacent to the streams at Odumante and these farms were highly intensive farms. The most dominant grass at the banks of the stream was *Panicum maximum*. The adjacent land-use at Abiriw was a riparian forest compared to that of Odumante which was predominantly farming. The stream flow in Odumante Sacred grove is a first-order or low-order (1 to 2) stream in the Strahler stream order system. These streams are classified as headwaters under the river continuum concept (Vannote et al., 1980; Doretto et al., 2020).

2.2. Study Area Characterization

2.2.1. Stream Size

The stream at Abiriw was deeper (30 - 50 cm in the rainy season and 15 - 45 cm in the dry season) compared to that of Odumante (10 - 30 cm in the rainy season and 2 - 10 cm in the dry season) (see [Table 2](#)).

2.2.2. Land Use within the Sacred Groves

The human impact within these sacred groves was assigned qualitative values as highly impacted, moderately impacted and less impacted. Areas of high intensive agriculture (use of pesticides and other chemicals) within these sacred groves, especially at Odumante were labelled as highly impacted, places of less intensive agriculture (no usage of agro-chemicals) were labelled as moderately impacted and semi natural forests where the inhabitants actively collected firewood were indicated as less impacted places. We randomly selected four sites, each within the two sacred groves based on the levels of impact as stated earlier. In our site selection we assigned two of the sites at Odumante as highly impacted and two other sites as moderately impacted areas and assigned all sites at Abiriw as less impacted areas.

2.3. Data Collection

Data collection was conducted via sampling different microhabitats associated with the different impacts (especially at Odumante) associated with these streams.

The sampled microhabitats were under cobbles, rocks, decaying leaves, sand and dislodgable boulders. There were four sampling sites per sacred grove (making eight sampling sites in total for the study) with each sampling site being sampled four times per season, making a total of eight sampling times per each sampling site for the entire study. The sampling period covered a one calendar year covering both dry and rainy seasons on four sampling events per each season per study site. We used Dip net and Surber sampler to sample for benthic insects. The Dip net was used to sample in the riffle areas, pool and edges of the stream. A handful of slightly decomposed and brownish leaf packs were collected with the dip nets. The benthic insects within these streams were also sampled with a Surber Stream-bottom sampler as described by Hynes (1975). Each of the streams was sampled on four occasions (replications) at four different sampling sites making a total of sixteen sampling events per stream per season (thirty-two in total per stream per year), for each of the different sampling methods described above. In total, there were sixty-four sampling events conducted across the two sacred groves for the study. We pooled the samples from the Dip net and Surber sampler before data analysis. We used the insect community analysis under NMDS to deduce the possible pollution level of the two streams.

2.4. Water Quality Analysis

The Eureka Sub3 multiparameter water-quality multiprobe was used to measure the physico-chemical parameters of the streams such as dissolved oxygen (DO), temperature, optically dissolved oxygen, conductivity, pH, and turbidity. The Eureka Sub3 multiparameter multiprobe is a portable water quality measuring device that is inserted directly into the water body. The device was calibrated prior to use for each sampling event for the four sampling occasions to ensure uniformity in measurement. The Eureka Sub3 multiparameter multiprobe was placed directly into the streams at the randomly selected points where each of the four sampling events occurred to record the DO, temperature, optical DO, conductivity, pH, and turbidity. The Eureka Sub3 multiparameter multiprobe is a multiple sensor holding single probe which can be used to measure several water quality parameters like: Oxidation-Reduction Potential (ORP), Temperature, Salinity/Conductivity, pH, Turbidity, Depth and Dissolved Oxygen (DO) at a goal and at the same location.

The advantage of using Eureka Sub3 multiparameter multiprobe water-quality multiprobe in sampling is that it gives a higher accurate and a more realistic information about the environmental factors being studied compared to measuring each factor separately. Samples from each of the streams were also taken to the laboratory for confirmatory analysis on the water quality parameters. There were two readings per each sampling event at each sampling site (one on the multiprobe and the other from confirmatory lab analysis). The average of the two readings were recorded as one reading per site per sampling event. In total, there were sixty-four readings comprising of four readings per sampling event on four occasions per season within the two seasons and used for the analysis. The average of these readings are summarized on **Table 1**.

Table 1. Diversity indices of aquatic insects of sacred groves during study period.

Diversity Indices	Odumante	Abirw
Shannon-Wiener Index (H')	3.41	3.04
Pielou's Index (E_H)	0.83	0.79
Jaccard's Index (C_j)	0.58	0.58

2.5. Sorting and Identification

The number of insects occurring in each family was counted and recorded. We identified all insects with the aid of identification keys developed by Scholtz & Holm (1985), Klotz et al., (2008), Samways (2008); Ball & Morris (2015), Gibb & Oseto (2019) to the lowest possible taxon.

2.6. Data Analysis

For each study area, Shannon-Wiener diversity index, $H' = -\sum_{i=1}^S p_i \ln(p_i)$ was estimated. Pielou index, $E_H = H/H_{\max}$ was estimated. Jaccard index of Similarity, $C_j = j/(a + b - j)$, was used to calculate the similarity between the two sacred groves. We also conducted a Non-Metric Multidimensional Scaling (NMDS) analysis on the total insect assemblage structure for the different levels of human impact within Abirw (all less impacted) and Odumante (highly to less impacted) Sacred groves using the SPSS statistical package.

We further measured the Bray Curtis similarities of the various insect assemblages from the different sampling sites from Analysis of Similarity (ANOSIM) in SPSS. We used ANOSIM because it is a robust non-parametric statistic for testing for resemblances among sampled groups. We then used sample paired t-test to determine if diversity and abundance of insects were significantly different between the two sampled streams and seasons. Because the data was count data, with positive values, right skewed and heteroscedastic, it was subjected to log transformation to stabilize the variances and render it close to normal as possible. Test for normality was then conducted through Shapiro-Wilk test for normality.

The analysis was conducted at 5% significance level. Sampled paired t-test was more appropriate to analyze the transformed values of the investigated paired samples (two seasons and two streams) which meant testing whether the geometric mean ratio between the values for the different seasons or the different streams was equal to 1. The demerits associated with using paired t-test are that the number of rare taxa in data can drive non-normal differences and increase skewness and changes in community composition may not be expressed. This was corrected as much as possible through data transformation although there were just a few rare species in the dataset. The values assigned to these study sites for impact or human activities were used to construct insects assemblage community structures. Data analysis was conducted using SPSS software. There were two readings for the water quality analysis conducted.

3. Results

3.1. Comparison of Insect Abundance in the Two Sacred Groves

The study recorded 2767 individual aquatic insects belonging to 65 families from 11 orders for both sacred groves. A total of 1307 individuals, representing 47.2% of the total collection was realized at Odumante and the rest (1460 individuals) representing 52.8% of the total was collected at Abiriw. There were 19 unique insect families collected at Odumante, 5 unique families collected at Abiriw only with the two sacred groves sharing 41 common families. The most diverse insect family recorded was the Diptera while the most abundant was Hemiptera (about 40% of the entire collection).

There was no significant difference in the number of specimens sampled from the two sacred groves ($t = 0.9, p > 0.05$). The results also revealed that Odumante recorded a significantly higher insect families (60) compared to that of Abiriw (46) ($t = 3.2, p < 0.05$) (Figure 1). Furthermore, there were 30 families and 356 specimens collected in the rainy season which was significantly lower than that of dry season with 57 families and 964 individuals at Odumante ($t = 3.6, p < 0.05$). At Abiriw, the total number of families and individuals collected were 29 and 306 respectively, during the rainy season which was significantly lower than that of dry season with 40 families and 1154 specimens ($t = 3.8, p < 0.05$) (see Figure 1).

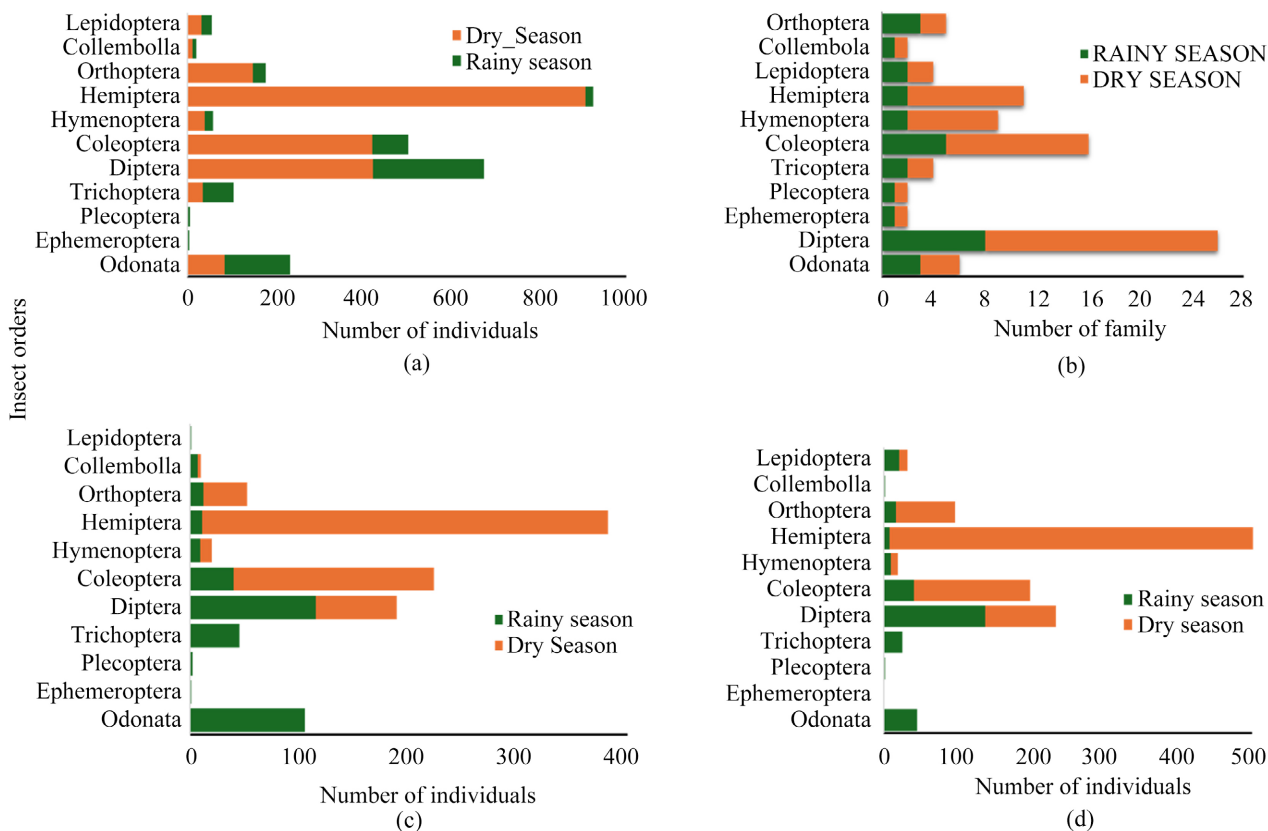


Figure 1. Graphs showing the total Insect abundance (a) and family richness (b) for the study and a comparison between the number of insects per season at the two study areas Odumante (c) and Abiriw (d) Sacred groves.

3.2. Abundance of Ephemeroptera, Plecoptera and Trichoptera (EPT)

The orders Ephemeroptera, Plecoptera and Trichoptera (EPT) recorded the least number of insects during the study. The percentage EPT in family diversity and abundance were 12.3% and 4.1%, respectively. The *Baetis* genus was the only Ephemeropteran genus sampled from the two sacred groves. Also, *Neoperla* was the only genus of Plecoptera sampled for the study while *Hydropsychidae* Sub-families *Diplectroninae* and *Hydropsychinae* were the sampled genus for the study. The most common Odonata genera sampled in these sacred groves were *Brachythemis*, *Crocothemis*, *Orthetrum*, *Trithemis*, and *Aethriamanta*.

3.3. Comparison of Insect Diversity between the Two Streams

3.3.1. Diversity Indices

The value of the diversity indices recorded per study site shows that diversity at Odumante was slightly higher with a value of 3.41 compared to that of Abiriw, which is 3.04, and with a Jaccard similarity index of 0.58 for both sacred groves. (Table 1).

Both sacred groves recorded an even distribution of insect species and in both seasons (Table 2).

Table 2. Diversity indices of both sacred groves in the wet and dry seasons.

Diversity Index	Wet Season		Dry Season	
	Odumante	Abiriw	Odumante	Abiriw
Shannon-Wiener Index (H')	2.71	2.69	2.87	2.48
Pielou's Index (E_H')	0.80	0.80	0.71	0.67

3.3.2. T-Test Results for Insect Diversity

The results of a student t-test between insect diversity of aquatic insects between the two streams were not significantly different ($t = 0.87$, $p > 0.05$) despite marginal differences observed during the study, however, diversity was significantly higher in the dry season compared to the rainy season within the two sacred groves ($t = 3.3$, $p < 0.05$).

3.3.3. Water Quality Analysis within the Two Streams

The water quality analysis results show that stream temperature remained same within the two study streams. However, there was a general increase in temperature during the dry season compared to the rainy season in both streams. Although turbidity in these streams was slightly higher during the dry season compared to rainy season, the levels were relatively low in both streams, indicating relatively clear water conditions. The most observable difference between the two streams in the two study areas was the amount of dissolved oxygen (DO) levels recorded. The DO level remained relatively same in both seasons in Abiriw indicating a stable and healthy oxygen ecosystem in contrast to that of Odumante

which showed a more pronounced reduction from the rainy season to the dry season with the average dissolved oxygen (DO) significantly reducing at Odumante during the dry season ($t = 3.10$, $p < 0.05$). The pH values and the amount of rock and sand substrates recorded were similar across both study areas and seasons. This shows stable chemical and habitat conditions for aquatic life in both streams. However, the riparian lands adjacent to the stream at Abiriw were surrounded by riparian forest while the stream at Odumante was surrounded by cultivated lands (See **Table 3**).

Table 3. Stream characteristics of the sacred groves within the study area.

Study Area	Abiriw		Odumante	
	Rainy	Dry	Rainy	Dry
Stream order	2	2	1	1
Stream depth (cm)	30 - 50	15 - 45	10 - 30	2 - 10
Stream width (cm)	135 - 567	125 - 367	108 - 189	108
Average Temperature (°C)	23	24.5	24	25.2
Average Turbidity (NTU)	3	4	3.3	3.5
Average Dissolved oxygen (mg/l)	7.5	7	6.5	4.5
Dominant substrates	Rock/Sand	Rock/Sand	Rock/Sand	Rock/Sand
Adjacent land use	Riparian forest	Riparian forest	Cultivated	Cultivated
Average pH	6.7	6.8	6.8	6.9

3.4. Insect Community Analysis

Overall, the total insect abundance and diversity were significantly higher in the dry season compared to the rainy season ($t = 4.1$, $p < 0.05$). See **Figure 1**.

The results of the Non-Metric Multidimensional Scaling (NMDS) analysis showed that the dispersion of insect assemblages between study sites was not very wide apart between sites. This is because there were no significant variations in insect assemblage communities between the different sites within all sacred groves ($R = 0.2$, $p < 0.001$). This could be seen by the nature of spread of the community structure where all the different impacted sites converged or clustered along the same plains on the NMDS graphs for Abiriw ($R = 0.02$, $p < 0.01$), indicating similarities in community structures although that of Odumante showed a bigger spread and differences ($R = 0.36$, $p < 0.01$) in sites (NMDS, stress = 0.07) (**Figure 2**).

4. Discussion

Our findings show the importance of environmental gradients, especially seasonality and stream dynamics in shaping aquatic insect communities within sacred groves. Although both streams examined in this study were subjected to varying

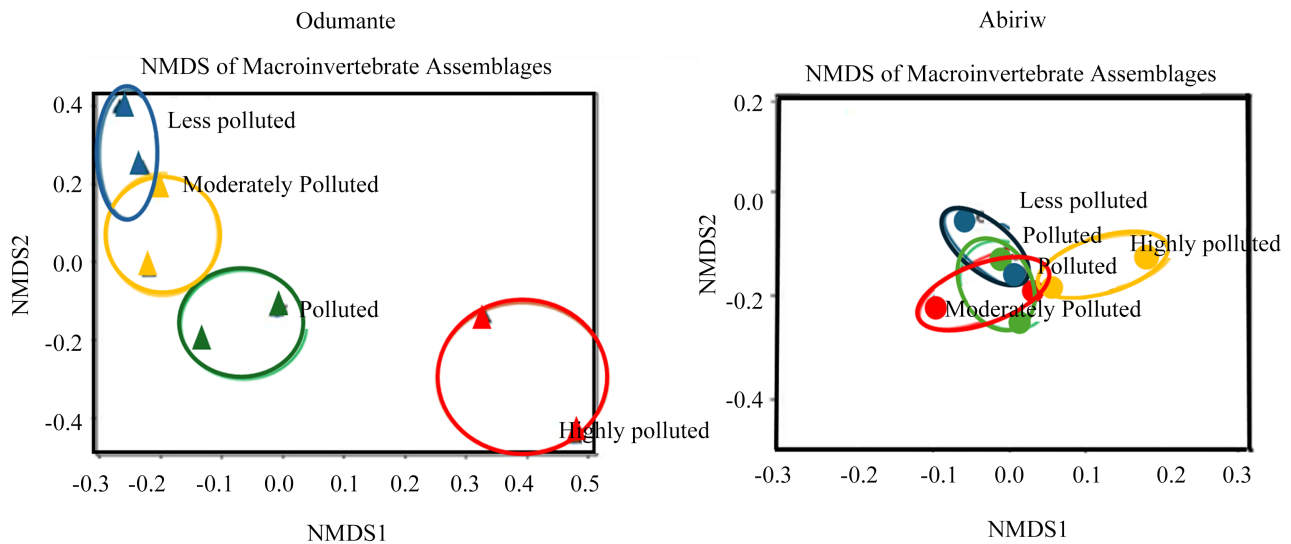


Figure 2. A Non-Metric Multidimensional Scaling (NMDS) ordination of the sampling sites (triangular and circular points) sampled within the study area, depicting insect assemblage structure within these sites. Sites categorized into same impacts fall within circles (NMDS, stress = 0.07).

degrees of impacts with Odumante more impacted compared to Abiriw, we inferred from the results that seasonal differences and stream dynamics were the main determining factors of aquatic invertebrate diversity within the sacred groves. Specifically, the high diversity recorded at Odumante shows that small streams with high to moderate anthropogenic impacts create an enabling environment for high insect diversity. This finding is consistent with published research (Stanford & Ward, 1983; Agra et al., 2021; Bouchon et al., 2025).

Seasonal shifts, especially the transition between rainy season and dry season, and periods of high temperature cause distinct alterations in aquatic insect assemblages, as reported elsewhere (Twining et al., 2019; Ergović et al., 2025; Ferreira et al., 2025; Kumari & Meena, 2026). These hydrological cycles drive predictable ecological rhythms, influencing oxygen levels, substrate composition, current velocity, and nutrient flux, which in turn affect the recruitment, emergence, and mortality of different insect taxa. Our observation of higher abundance and richness in the dry season aligns with studies showing that low-flow periods offer stable microhabitats conducive to insect aggregation, predator proliferation, and resource retention (Kasangaki et al., 2008; Basumatary, 2023).

Insect diversity may differ from one stream to another, probably in relation to differences in flow, oxygen concentration and other factors such as the accumulation of organic debris (Vannote et al., 1980; Birrell et al., 2020; Doretto et al., 2020; Findlay, 2021). Rainfall affects water levels and current strengths of aquatic insect habitats. For instance, net-spinning caddisflies and stoneflies suffer population reduction during heavy rains that could cause floods (Jennings et al., 2023; Phillips et al., 2023). The significantly higher insect abundance and diversity observed in the dry season support patterns observed in other tropical freshwater systems. Reduced flow during the dry season creates stable microhabitats that pro-

mote aggregation of aquatic insects, particularly collectors and predators. Also, the significantly lower DO levels in these streams may be an indication of oxygen stress affecting breeding sites in these streams during dry seasons although there is physical stability during same period (Simaika & Macadam, 2024; Enns et al., 2025; Jaulin, 2025). In contrast, the rainy season often brings heavy flow variability and turbidity, leading to displacement and reduced capture efficiency. These seasonal shifts influence beta diversity and community composition, a trend observed in similar studies elsewhere (Kasangaki et al., 2008; Sánchez-Argüello et al., 2010; Dong et al., 2021; Tan et al., 2021; Wang et al., 2023; Yang et al., 2025).

We could draw direct inferences from our results that the increase in temperature of the streams during dry season could be because of the loss of shade cover coupled with the high solar radiation in the study areas, as reported elsewhere (Johnson, 2004; Windoloski, 2024). Also, the little differences in the turbidity of the two sacred groves and in the different seasons may be an indication of minor anthropogenic disturbance or a reduction in water volumes especially in the dry season (Windoloski, 2024; Morilla et al., 2026). Furthermore, the decline in DO in the streams, especially in the dry season, is an indication of environmental stress which can adversely affect aquatic insects (Omokunle, 2024; Nguyen & Nguyen, 2025). Further, the pH values recorded for this study show stable chemical and habitat conditions for aquatic life in both streams (Omokunle, 2024; Nguyen & Nguyen, 2025). Also, the largely secondary forests at the riparian areas of the Abirw stream could have ensured relatively better water quality in that stream by offering protection from erosion through better shading, and a better natural filtration system (Vero & Snell, 2025; Morilla et al., 2026). In contrast to Abirw sacred grove, the Oduamante stream with highly intensive riparian agricultural activities can potentially contribute to increased water and nutrient runoff into the stream and thereby affecting the water quality as was observed in the reduced dissolved oxygen levels (Vero & Snell, 2025; Morilla et al., 2026).

The dominance of Hemiptera is an indicator of low levels of dissolved oxygen, moderate pollution, high nutrients and some amount of habitat disturbance in these streams especially within the Odumate sacred grove. This is because Hemiptera is known to tolerate oxygen stressed aquatic environment better than most EPT taxa. This is due to their characteristic carriage of air bubbles under water and constant respiration at the water surface. Also, several studies have shown that Hemiptera are usually tolerant in moderately polluted environment (Bakonyi et al., 2022; El Yaagoubi et al., 2024; Omokunle, 2024; Wani et al., 2024).

The results of insect diversity during the rainy season contrast with other studies that showed that aquatic insect abundance in the rainy season is relatively higher compared to dry season in streams and rivers (Santana et al., 2015; Yuen & Dudgeon, 2016; Pedrosa Guimarães et al., 2025) but in support with results of other studies (Ferreira et al., 2025; Zhang et al., 2025). This may be because most aquatic insects in the study area might have been displaced from their habitats due to increased flow rate and strong current that normally accompany rainfall in the

study area as has been reported in earlier studies (Godoy et al., 2022; Cooke et al., 2022; Marques et al., 2025). Stream dynamics critically moderates insect community structure by mediating trade-offs between taxonomic richness and abundance (Vannote et al., 1980; Kasangaki et al., 2008; Lehti, 2025). The NMDS for the two study areas were clearly distinct, especially in Odumante, supporting this analysis as was the result in other studies elsewhere (Susanto et al., 2023; Zhao et al., 2023).

Furthermore, the smaller Odumante stream exhibited higher family richness despite lower abundance, contrasting sharply with the larger Abiriv stream where greater water volume supported higher abundance but reduced diversity. This result probably emanates from environmental filtering processes at Abiriv, with an extensive catchment area, amplified diffuse pollution and sedimentation, reducing microhabitat heterogeneity which favours pollution-tolerant generalists (Brasil et al., 2020; Cararo et al., 2026); whereas at Odumante, constrained dimensions maintained heterogeneous microhabitats that buffered localized disturbances, enabling specialized guild persistence (Sankone et al., 2023). Consequently, stream dynamics driven environmental heterogeneity increased directly might have shaped community outcomes explaining Odumante's relatively higher richness and Shannon diversity versus Abiriv's abundance-dominated assemblages. The river continuum concept which states that small streams such as the current study streams (1 - 3 Strahler stream order) are classified as headwaters which are heavily influenced by riparian vegetations by leaving allochthonous detritus and its shading effect (Vannote et al., 1980; Doretto et al., 2020) could also be used to explain this dynamic of high diversity at Odumante.

The riparian vegetation at Odumante sacred grove was found to have been highly impacted by human activities which have eventually affected the physico-chemical characteristics of the water specifically the reduction of DO but might have led to increased allochthonous detritus and thereby providing more opportunities and support for higher diversity. This nonetheless was at the expense of high abundance which may be due to low oxygen levels that will eventually impact breeding sites and increased inter and intra-specific competition, leading to a reduction in individual populations and other such impacts especially during the dry season and may be a sign of relatively some level of pollution which ends up affecting the most sensitive groups like the EPT groups. This demonstrates how intrinsic stream dynamics and seasonal influence community structure in freshwater ecosystems.

The major aquatic insect orders of Ephemeroptera, Plecoptera and Trichoptera which are very sensitive to pollutants were poorly represented at both sacred groves in both seasons. This may be an indication of the poor water quality of the streams, including oxygen stress as reported by (Akamagwuna et al., 2022; Tubic et al., 2024; Barasa et al., 2025). Also, the dominance of Diptera, Hemiptera and Coleoptera in both sacred groves, was a confirmation that these insect orders were the most described and largest in relations to other insect orders and they can

inhabit almost every habitat in the aquatic environment in this case in the two sacred groves in agreement with other studies (Amusan & Akin-Aina, 2024; Liebhold et al., 2024; Akinmuleya & Adu, 2025).

Furthermore, this study showed that the persistence of Odonata especially Libellulidae on the aquatic landscape was heavily influenced by structures within the environment, mostly biosphere such as shade cover (Santos & Rodrigues, 2022; Worthen & Guevara-Mora, 2024; Kaltsas et al., 2025) and land use type (Rodriguez-Tapia et al., 2022; Calvão et al., 2023) and micro-climate (Marta et al., 2021; Guim Ursul et al., 2024). Acquah-Lampitey et al., (2013), Dijkstra & Clausnitzer (2014), and De Knijf & Van Grunsven (2015), point out that the degree of shadiness seems to be a principal cue for Odonates to select a forest habitat, but the forest type (species composition) as well as the age of the forest bordering running water sites is of little influence for the Odonata assemblages as long as the required cover is present. These conditions were normally depicted in Abiriw.

Studies have shown that EPT are very sensitive to pollution within aquatic ecosystems. For instance, studies have reported that they are very sensitive to sediment, heavy metals, eutrophication pesticides, etc. pollution (Adubor et al., 2025; Barasa et al., 2025) within the aquatic ecosystem. In general, these studies have shown that low diversity and abundance of EPT is an indication of pollution by high nutrient levels or reduced dissolved oxygen in the water body. The result of our study aligns with these findings and further confirmation that these streams might be polluted or have low dissolved oxygen levels. The low levels of EPT may also be due to chemical runoff from nearby agricultural fields, and the loss of forest cover and land-use change in the riparian environment especially at Odumante as reported in Adubor et al. (2025), Barasa et al., 2025 and El Yaagoubi et al., (2025). The low levels of EPT coupled with the dominance of Diptera in our studies are an indicator of declining macroinvertebrate diversity within these streams (Tubić et al., 2024; Barasa et al., 2025; Indraswari Suhri et al., 2025).

5. Conclusion

Overall, the study demonstrates that seasonal variation, spatial location, and stream dynamics specifically environmental stress like low oxygen in the stream especially during dry season and pollution from agricultural fields significantly influence aquatic insect diversity and abundance in the study areas especially at Odumante sacred grove. These findings underscore the ecological importance of preserving both sites, especially during the dry season when aquatic insect communities are more diverse and abundant. The study further showed that both Odumante and Abiriw sacred groves harbour a high number of aquatic insects. The traditional norms and taboos which were previously used to deter people from encroaching in the sacred groves appeared to have been relaxed, resulting in encroachment and pollution of the streams.

Odumante sacred grove had a higher insect diversity compared to Abiriw sacred grove even though Odumante was more fragmented. One major contribu-

tory factor to the low insect diversity at Odumante was human impact stemming from the surrounding villages along the upper reaches of the stream to the sacred grove. It is also concluded from this study that aquatic insects prefer impacted environment especially for streams in sacred groves classified under first-order or low-order (1 to 2) in the Strahler stream order system because according to the river continuum concept such streams are classified as headwaters and are heavily influenced by riparian vegetations by increasing shading and allochthonous detritus which further gives rise to more resources and increased heterogenous environment. The low abundance of EPT shows that the stream dynamics such as dissolve oxygen is low and pollution from runoff from agricultural fields is increasing. The EPT diversity in this study can therefore be used as surrogate organisms to conclude on the stream dynamics that the stream is moderately to severely impacted. We recommend that there should be reduced human activities to slow human impact in these streams in these sacred groves to improve aquatic insect biodiversity conservation.

Acknowledgements

The authors would like to express their most sincere thanks to the Department of Animal of Animal Biology and Conservation Science of University of Ghana for their support for this research. The authors would also like to acknowledge the leadership and local community for granting them access to the two sacred groves.

Availability of Data

Data will be made available on request.

Funding

This research was self-funded by the authors.

Authors' Contributions

Victor Parry: Conceptualization, Writing of manuscript, Data collection and analysis; Roger, Sigismund Andersen: Conceptualization, Data collection and analysis and identification of insect groups, Samuel Adu-Acheampong: Data processing, Writing of manuscript, Review & editing of manuscript, Mukundi Mukundamago: Writing of manuscript, Review & editing of manuscript, Emmanuel Robert Blankson: Writing of manuscript, Review & editing of manuscript, Rosina Kyerematen; Conceptualization, Writing review & editing of manuscript and Supervision.

Conflicts of Interest

The authors declare that they have no conflict of interest.

References

Acquah-Lampsey, D., Kyerematen, R., & Owusu, E. O. (2013). Dragonflies (Odonata: An-

- isoptera) as Tools for Habitat Quality Assessment and Monitoring. *Journal of Applied Biosciences Research*, 2, 178-182.
- Adarkwa-Dadzie, A. (1997). *The Contribution of Ghanaian Beliefs to Biodiversity Conservation*. Paper Presentation, UNESCO-MAB Seminar on Biosphere Reserves for Biodiversity Conservation and Sustainable Development in Anglophone Africa (BRAAF), Cape Coast. Ghana.
- Adler, P. H., & Footitt, R. G. (2009). *Insect Biodiversity: Science and Society*. Blackwell Publishing.
- Adu-Acheampong, S., & Samways, M. J. (2019). Traits and Land Transformation Change the Fortunes of Grasshopper Generalists vs. Specialists in a Biodiversity Hotspot. *Bio-systems Diversity*, 27, 26-32. <https://doi.org/10.15421/011904>
- Adu-Acheampong, S., Bazelet, C. S., & Samways, M. J. (2016). Extent to Which an Agricultural Mosaic Supports Endemic Species-Rich Grasshopper Assemblages in the Cape Floristic Region Biodiversity Hotspot. *Agriculture, Ecosystems & Environment*, 227, 52-60. <https://doi.org/10.1016/j.agee.2016.04.019>
- Adubor, C., Ekperusi, A. O., Michael, A., & Olomukoro, J. O. (2025). Physicochemical Properties of Surface Water, Heavy Metals Levels in Sediments and Macrobenthic Invertebrates Community of Ikpoba River, Benin City, Edo State, Nigeria. *Journal of Applied Sciences and Environmental Management*, 29, 1653-1663. <https://doi.org/10.4314/jasem.v29i5.33>
- Agodzo, S. K., Bessah, E., & Nyatuame, M. (2023). A Review of the Water Resources of Ghana in a Changing Climate and Anthropogenic Stresses. *Frontiers in Water*, 4, Article ID: 973825. <https://doi.org/10.3389/frwa.2022.973825>
- Agra, J., Ligeiro, R., Heino, J., Macedo, D. R., Castro, D. M. P., Linares, M. S. et al. (2021). Anthropogenic Disturbances Alter the Relationships between Environmental Heterogeneity and Biodiversity of Stream Insects. *Ecological Indicators*, 121, Article 107079. <https://doi.org/10.1016/j.ecolind.2020.107079>
- Akamagwuna, F. C., Edegbene, A. O., Ntloko, P., Arimoro, F. O., Nnadozie, C. F., Choruma, D. J. et al. (2022). Functional Groups of Afrotropical EPT (Ephemeroptera, Plecoptera and Trichoptera) as Bioindicators of Semi-Urban Pollution in the Tsitsa River Catchment, Eastern Cape, South Africa. *PeerJ*, 10, e13970. <https://doi.org/10.7717/peerj.13970>
- Akinmuleya, C. B., & Adu, F. D. (2025). Effects of Anthropogenic Activities on Selected Insect Species Orders (Hymenoptera, Orthoptera and Hemiptera) in Afuremo, Oye-Egbo and Faalex Areas of Oye-Ekiti, Ekiti State, Nigeria. *Journal of Applied Sciences and Environmental Management*, 29, 2013-2020. <https://doi.org/10.4314/jasem.v29i6.35>
- Amusan, B. O., & Akin-Aina, O. F. (2024). Diversity and Seasonal Abundance of Aquatic Insects Associated with the Littoral Zone of a Tropical Inland Water. *Journal of Life Science*, 4, 14-21.
- Anamika, R., Kumari, V., & Meena, S. (2026). Seasonal Dynamics and Biomonitoring Role of Aquatic Entomofauna in an Indian Artificial Lake. *Environmental Science and Pollution Research*, 33, 2303-2318. <https://doi.org/10.1007/s11356-025-37379-z>
- Anderson, R. S., Billah, M. K., Acquah-Lampsey, D., Aidoo, O. F., Adu-Acheampong, S., Gyimah, T. et al. (2025). Rapid Assessment of Insect Responses Post-Fire Chronosequence in Equal-Sized Savanna Grassland Plots. *African Journal of Ecology*, 63, e70094. <https://doi.org/10.1111/aje.70094>
- Aniah, P., & Yelfaanibe, A. (2018). Environment, Development and Sustainability of Local Practices in the Sacred Groves and Shrines in Bongo District: A Bio-Cultural Study for Environmental Management in Ghana. *Environment, Development and Sustainability*, 20, 2487-2499. <https://doi.org/10.1007/s10668-017-0001-2>

- Anokye, F. A. (2022) *Indigenous Knowledge and Coastal Livelihood Resource Sustainability at Moree and Apam Communities in the Central Region of Ghana*. Doctoral Thesis, University of Cape Coast.
- Asante, D. B. (2024). Exploring the Spiritual and Functional Aspects of Sacred Groves: An Ecological Case Study of the Kwaékese and Twendurase Mystical Sacred Groves in Ghana. *Worldviews: Global Religions, Culture, and Ecology*, 28, 246-267. <https://doi.org/10.1163/15685357-02803003>
- Asokan, A., Chouhan, S., & Singh, V. (2015). Sacred Grove—A Nature’s Gift—As a Remedy for Human Ailments, a Biodiversity Reservoir for Restoring Indigenous Traits for Endangered Listed Plants—A Review. *Open Access Library Journal*, 2, 1-13. <https://doi.org/10.4236/oalib.1101517>
- Attuquayefio, D., & Folib, J. (2005). An Overview of Biodiversity Conservation in Ghana: Challenges and Prospects. *West African Journal of Applied Ecology*, 7, 1-18. <https://doi.org/10.4314/wajae.v7i1.45621>
- Ayivie, D. (2005) *Description of Three New Species of Chironominae Macquart and a New Orthoclaadiinae Genus from the Abiriw Sacred Grove, Eastern Region of Ghana*. Master’s Thesis, University of Ghana.
- Bakonyi, G., Vászrhelyi, T., & Szabó, B. (2022). Pollution Impacts on Water Bugs (Nepomorpha, Gerromorpha): State of the Art and Their Biomonitoring Potential. *Environmental Monitoring and Assessment*, 194, Article No. 301. <https://doi.org/10.1007/s10661-022-09961-2>
- Ball, S., & Morris, R. (2015). *Britain’s Hoverflies: A Field Guide*. Princeton University Press.
- Barasa, C. N., Simiyu Muse, G., & Khazenzi, J. A. (2025). Human Activities Influence on Distribution of Ephemeroptera, Plecoptera, and Trichoptera (EPT) in a Tropical Riverine Ecosystem in Kenya. *Journal of Freshwater Ecology*, 40, Article 2539125. <https://doi.org/10.1080/02705060.2025.2539125>
- Basumatary, S. (2023). The Diversity of Aquatic Insects in Assam, North-East India: A Review. *International Journal of Biosciences*, 23, 230-240.
- Batame, M., Sarfo, I., Yeboah, E., Njomaba, E., & Pupilampu, D. A. (2023). Mapping of Sacred Groves in Ghana: The Case of Talensi District in the Guinea Ecological Zone. *SN Social Sciences*, 3, Article No. 145. <https://doi.org/10.1007/s43545-023-00737-0>
- Birrell, J. H., Shah, A. A., Hotaling, S., Giersch, J. J., Williamson, C. E., Jacobsen, D. et al. (2020). Insects in High-Elevation Streams: Life in Extreme Environments Imperiled by Climate Change. *Global Change Biology*, 26, 6667-6684. <https://doi.org/10.1111/gcb.15356>
- Boamah, R. (2020). *An Assessment of Effects of Illegal Activities on Timber Production in Southern Scarp Forest Reserve*. Doctoral Dissertation, University of Cape Coast.
- Bonada, N., Prat, N., Resh, V. H., & Statzner, B. (2006). Developments in Aquatic Insect Biomonitoring: A Comparative Analysis of Recent Approaches. *Annual Review of Entomology*, 51, 495-523. <https://doi.org/10.1146/annurev.ento.51.110104.151124>
- Bouchon, Y. J., Quinteiro, F. B., & Beasley, C. R. (2025). *Aquatic Insects in a Forested and an Urban Stream: The Importance of Riparian Forest, Microhabitats and Climate Influence*. <https://doi.org/10.21203/rs.3.rs-8029671/v2>
- Brasil, L. S., de Lima, E. L., Spigoloni, Z. A., Ribeiro-Brasil, D. R. G., & Juen, L. (2020). The Habitat Integrity Index and Aquatic Insect Communities in Tropical Streams: A Meta-Analysis. *Ecological Indicators*, 116, Article 106495. <https://doi.org/10.1016/j.ecolind.2020.106495>
- Calvão, L. B., Brito, J. d. S., Ferreira, D., Cunha, E. J., Oliveira-Junior, J. M. B., & Juen, L.

- (2023). Effects of the Loss of Forest Cover on Odonate Communities in Eastern Amazonia. *Journal of Insect Conservation*, 27, 205-218. <https://doi.org/10.1007/s10841-022-00444-w>
- Cararo, E. R., Bini, L. M., & Johansson, F. (2026). Beyond Pond Size: The Roles of Disproportionate Effects and Habitat Heterogeneity in Controlling Biodiversity in Urban Ponds. *Urban Ecosystems*, 29, Article No. 11. <https://doi.org/10.1007/s11252-025-01878-1>
- Carvalho, R. L., Andersen, A. N., Anjos, D. V., Pacheco, R., Chagas, L., & Vasconcelos, H. L. (2020). Understanding What Bioindicators Are Actually Indicating: Linking Disturbance Responses to Ecological Traits of Dung Beetles and Ants. *Ecological Indicators*, 108, Article 105764. <https://doi.org/10.1016/j.ecolind.2019.105764>
- Chowdhury, S., Dubey, V. K., Choudhury, S., Das, A., Jeengar, D., Sujatha, B. et al. (2023). Insects as Bioindicator: A Hidden Gem for Environmental Monitoring. *Frontiers in Environmental Science*, 11, Article ID: 1146052. <https://doi.org/10.3389/fenvs.2023.1146052>
- Cooke, S. J., Galassi, D. M. P., Gillanders, B. M., Landsman, S. J., Hammerschlag, N., Gallagher, A. J. et al. (2022). Consequences of “Natural” Disasters on Aquatic Life and Habitats. *Environmental Reviews*, 31, 122-140. <https://doi.org/10.1139/er-2022-0050>
- Datto-Liberato, F. H., Lopez, V. M., Quinaia, T., do Valle Junior, R. F., Samways, M. J., Juen, L. et al. (2024). Total Environment Sentinels: Dragonflies as Ambivalent/Amphibiotic Bioindicators of Damage to Soil and Freshwater. *Science of The Total Environment*, 934, Article 173110. <https://doi.org/10.1016/j.scitotenv.2024.173110>
- De Knijf, G., & Van Grunsven, R. (2015). Recensies: The Dragonflies and Damsel-Flies of Eastern Africa. Handbook for all Odonata from Sudan to Zimbabwe. Klaas-Douwe B. Dijkstra and Viola Clausnitzer (2014). *Brachytron*, 17, 58-59.
- de Souza, K. A. (2025). *Relationships between Sacred Natural Sites, Ecological Corridors, and Priority Resilience Climate Change Areas in the Upper Paraguay River Basin*. Master's Thesis, Federal University of Mato Grosso do Sul.
- Diawuo, F., & Issifu, A. K. (2017). Exploring the African Traditional Belief Systems (Totems and Taboos) in Natural Resources Conservation and Management in Ghana. In J. Chimakonam (Ed.), *African Philosophy and Environmental Conservation* (pp. 209-221). Routledge. <https://doi.org/10.4324/9781315099491-16>
- Dijkstra, K. D. B., & Clausnitzer, V. (2014). *The Dragonflies and Damselflies of East-ern Africa: Handbook for All Odonata from Sudan to Zimbabwe*. African Dragonfly Fund.
- Doffana, Z. D. (2017). Sacred Natural Sites, Herbal Medicine, Medicinal Plants and Their Conservation in Sidama, Ethiopia. *Cogent Food & Agriculture*, 3, Article 1365399. <https://doi.org/10.1080/23311932.2017.1365399>
- Dong, R., Wang, Y., Lu, C., Lei, G., & Wen, L. (2021). The Seasonality of Macroinvertebrate B Diversity along the Gradient of Hydrological Connectivity in a Dynamic River-Floodplain System. *Ecological Indicators*, 121, Article 107112. <https://doi.org/10.1016/j.ecolind.2020.107112>
- Doretto, A., Piano, E., & Larson, C. E. (2020). The River Continuum Concept: Lessons from the Past and Perspectives for the Future. *Canadian Journal of Fisheries and Aquatic Sciences*, 77, 1853-1864. <https://doi.org/10.1139/cjfas-2020-0039>
- El Yaagoubi, S., Edegbene, A. O., El Haissoufi, M., Harrak, R., & El Alami, M. (2024). Odonata, Coleoptera, and Heteroptera (OCH) Trait-Based Biomonitoring of Rivers within the Northwestern Rif of Morocco: Exploring the Responses of Traits to Prevailing Environmental Gradients. *Ecologies*, 5, 132-154. <https://doi.org/10.3390/ecologies5010009>
- El Yaagoubi, S., Errochdi, S., Edegbene, A. O., Kassout, J., Harrak, R., & El Alami, M.

- (2025). Distribution Patterns of Ephemeroptera, Plecoptera, and Trichoptera (EPT) Species in the Northwestern Rif: Environmental and Climate Change Impacts. *Hydrobiologia*, 852, 1575-1593. <https://doi.org/10.1007/s10750-024-05756-3>
- Enns, D., Baker, N. J., Oehlmann, J., & Jourdan, J. (2025). Stream Baseline Conditions Shape Functional Responses to Wastewater: Evidence from Insect-Dominated Sites. *PeerJ*, 13, e20193. <https://doi.org/10.7717/peerj.20193>
- Ergović, V., Čerba, D., Tubić, B., Novaković, B., Koh, M., & Mihaljević, Z. (2025). Seasonal Dynamics and Factors Shaping Aquatic Insect Assemblages in Mountain Streams of the Pannonian Lowland Ecoregion. *Insects*, 16, Article 344. <https://doi.org/10.3390/insects16040344>
- Ferreira, P. H. S., Lima, E. L. d., Lima-Junior, D. P., & Brasil, L. S. (2025). Climatic Seasonality of the Cerrado and Aquatic Insect Communities: A Systematic Review with Meta-Analysis. *Environmental Monitoring and Assessment*, 197, Article No. 358. <https://doi.org/10.1007/s10661-025-13783-3>
- Findlay, S. E. G. (2021). Organic Matter Decomposition. In K. C. Weathers, D. L. Strayer, & G. E. Likens (Eds.), *Fundamentals of Ecosystem Science* (pp. 81-102). Elsevier. <https://doi.org/10.1016/b978-0-12-812762-9.00004-6>
- Franks, P., Hou-Jones, X., Fikreyesus, D., Sintayehu, M., Mamuye, S., Danso, E. Y., Meshack, C. K., McNicol, I. & van Soesbergen, A. (2017). *Reconciling Forest Conservation with Food Production in Sub-Saharan Africa*. International Institute for Environment and Development.
- Gibb, T. J., & Oseto, C. (2019). *Insect Collection and Identification: Techniques for the Field and Laboratory*. Academic Press.
- Godoy, B. S., Valente-Neto, F., Queiroz, L. L., Holanda, L. F. R., Roque, F. O., Lodi, S. et al. (2022). Structuring Functional Groups of Aquatic Insects along the Resistance/Resilience Axis When Facing Water Flow Changes. *Ecology and Evolution*, 12, e8749. <https://doi.org/10.1002/ece3.8749>
- Gordon, C. (1992). *Sacred Groves and Conservation in Ghana* (Vol. 1, pp. 3-4). Newsletter of the IUCN SSC African Reptile and Amphibian Specialist Group.
- Guim Ursul, M., Juan Pablo Cancela, H., & Wilson, R. (2024). Chapter 16 Refugia from Climate Change and Their Influence on the Diversity and Conservation of Insects. In: D. González-Tokman and W. Dáttilo (Eds.), *Effects of Climate Change on Insects: Physiological, Evolutionary, and Ecological Responses* (pp. 329-352). Oxford. <https://doi.org/10.1093/oso/9780192864161.003.0016>
- Gyedu, A. K., Dampfey, F. G., Barnes, V. R., Bentsi-Enchill, F., Sarfo, D. A., Mohammed, I. et al. (2025). Multiple Anthropogenic and Climatic Factors Drive Tree Species Attributes in Ecologically Distinct Sacred Groves in Ghana. *International Journal of Forestry Research*, 2025, Article 9944297. <https://doi.org/10.1155/ijfr/9944297>
- Hodkinson, I. D., & Jackson, J. K. (2005). Terrestrial and Aquatic Invertebrates as Bioindicators for Environmental Monitoring, with Particular Reference to Mountain Ecosystems. *Environmental Management*, 35, 649-666. <https://doi.org/10.1007/s00267-004-0211-x>
- Hynes, J. D. (1975). Annual Cycles of Macro-Invertebrates of a River in Southern Ghana. *Freshwater Biology*, 5, 71-83. <https://doi.org/10.1111/j.1365-2427.1975.tb00122.x>
- Ikenyei, S. N., & Lawal, H. M. (2019) Forest and Social Control in Sapele, Okpe Kingdom, Delta State, Nigeria. *Lapai International Journal of Management and Social Sciences*, 11, 154-167.
- Ismail, N., Awg Abdul Rahman, A., Mohamed, M., Abu Bakar, M. F., & Tokiman, L. (2020).

- Butterfly as Bioindicator for Development of Conservation Areas in Bukit Reban Kambing, Bukit Belading and Bukit Tukau, Johor, Malaysia. *Biodiversitas Journal of Biological Diversity*, 21, 334-344. <https://doi.org/10.13057/biodiv/d210141>
- Jaulin, E. (2025). *Looking at the Reproduction of Stream Insects to Improve Our Understanding of the Ecological Effects of Artificial Flow Variations*. Doctoral Dissertation. Université Claude Bernard Lyon I.
- Jennings, C. A., Anderson, E. P., Benke, A. C., Kwak, T. J., Scott, M. C., & Smock, L. A. (2023). Atlantic Coast Rivers of the Southeastern United States. In M. D. Delong, T. D. Jardine et al. (Eds.), *Rivers of North America* (pp. 68-123). Elsevier. <https://doi.org/10.1016/b978-0-12-818847-7.00024-0>
- Johnson, S. L. (2004). Factors Influencing Stream Temperatures in Small Streams: Substrate Effects and a Shading Experiment. *Canadian Journal of Fisheries and Aquatic Sciences*, 61, 913-923. <https://doi.org/10.1139/f04-040>
- Kaltsas, D., Alvanou, L., Ekklesiarchos, I., Raptis, D. I., & Avtzis, D. N. (2025). Canopy Cover Drives Odonata Diversity and Conservation Prioritization in the Protected Wetland Complex of Thermaikos Gulf (Greece). *Forests*, 16, Article 1181. <https://doi.org/10.3390/f16071181>
- Karpiński, L., Maák, I., & Wegierek, P. (2021). The Role of Nature Reserves in Preserving Saproxyllic Biodiversity: Using Longhorn Beetles (Coleoptera: Cerambycidae) as Bioindicators. *The European Zoological Journal*, 88, 487-504. <https://doi.org/10.1080/24750263.2021.1900427>
- Kasangaki, A., Chapman, L. J., & Balirwa, J. (2008). Land Use and the Ecology of Benthic Macroinvertebrate Assemblages of High-Altitude Rainforest Streams in Uganda. *Freshwater Biology*, 53, 681-697. <https://doi.org/10.1111/j.1365-2427.2007.01925.x>
- Khan, S. R., Rastogi, N., & Singh, S. K. (2023). Bio-Transfer and Bioaccumulation Patterns of Heavy Metals in Mine Site-Inhabiting Ants and Grasshoppers, across Mine Site Restoration Chronosequence. *Ecotoxicology*, 32, 683-698. <https://doi.org/10.1007/s10646-023-02676-1>
- Klotz, J. H., Hansen, L. D., Pospischil, R., & Rust, M. (2008) *Urban Ants of North America and Europe: Identification, Biology and Management*. Cornell University Press.
- Kofi Sarfo-Adu, G., Kwabena Kokofu, H., Aferdi Dadebo, M., Nkrumah, G., & Kwaku Galle, D. (2022). Management of Sacred Groves and Customary Practices in Pursuit of Sustainable Forest Management. *Journal of Environment and Ecology*, 13, Article 31. <https://doi.org/10.5296/jee.v13i1.19836>
- Kosoe, E. A., Adjei, P. O., & Diawuo, F. (2020). From Sacrilege to Sustainability: The Role of Indigenous Knowledge Systems in Biodiversity Conservation in the Upper West Region of Ghana. *GeoJournal*, 85, 1057-1074. <https://doi.org/10.1007/s10708-019-10010-8>
- Kujur, R. S., Munda, S. S., Kumar, A., Rathna, V., & Lall, R. R. (2025). Sacred Grove: A Reservoir of Plant Diversity and Conservation. In R. K. Kalia, & R. Pathak (Eds.), *Tree Biology and Biotechnology* (pp. 133-144). Springer. https://doi.org/10.1007/978-981-96-0002-1_9
- Kumar, R., Prajapati, U., & Koli, V. K. (2022). Factors Driving the Tree Species Richness in Sacred Groves in Indian Subcontinent: A Review. *Biodiversity and Conservation*, 31, 2927-2943. <https://doi.org/10.1007/s10531-022-02474-x>
- Kwarteng, K. O. (2015). A History of Pre-Colonial and Colonial Wildlife Conservation Practices in Ghana. In M. Muchie & D. Millin (Eds.), *Between Rhetoric and Reality* (pp. 131-166). Langaa RPCIG. <https://doi.org/10.2307/j.ctvh9vwc4.8>
- Kyerematen, R., & Gordon, C. (2012). Aquatic Insect Fauna of three River Systems in the

- Akyem Abuakwa Traditional Area of the Eastern Region of Ghana. *Journal of Insect Science*, 20, 73-82.
- Kyerematen, R., Adu-Acheampong, S., Acquah-Lamprey, D., Anderson, R. S., Owusu, E. H., & Mantey, J. (2018a). Butterfly Diversity: An Indicator for Environmental Health within Tarkwa Gold Mine, Ghana. *Environment and Natural Resources Research*, 8, 69-83. <https://doi.org/10.5539/enrr.v8n3p69>
- Kyerematen, R., Kaiwa, F., Acquah-Lamprey, D., Adu-Acheampong, S., & Andersen, R. S. (2018b). Butterfly Assemblages of Two Wetlands: Response of Biodiversity to Different Environmental Stressors in Sierra Leone. *Open Journal of Ecology*, 8, 379-395. <https://doi.org/10.4236/oje.2018.87023>
- Lehti, N. (2025). *Effects of Habitat Management on Macroinvertebrate Community Structure and Diversity in Urban Aquatic Nature-Based Solutions*. Master's Thesis, University of Helsinki.
- Liebhold, A. M., Turner, R. M., Bartlett, C. R., Bertelsmeier, C., Blake, R. E., Brockerhoff, E. G. et al. (2024). Why So Many Hemiptera Invasions? *Diversity and Distributions*, 30, e13911. <https://doi.org/10.1111/ddi.13911>
- Lutinski, J. A., Lutinski, C. J., Serena, A. B., Busato, M. A., & Mello Garcia, F. R. (2024). Ants as Bioindicators of Habitat Conservation in a Conservation Area of the Atlantic Forest Biome. *Sociobiology*, 71, e9152. <https://doi.org/10.13102/sociobiology.v71i1.9152>
- Makwela, M. M., Slotow, R., & Munyai, T. C. (2023). Carabid Beetles (Coleoptera) as Indicators of Sustainability in Agroecosystems: A Systematic Review. *Sustainability*, 15, Article 3936. <https://doi.org/10.3390/su15053936>
- Maloney, K. O., Munguia, P., & Mitchell, R. M. (2011). Anthropogenic Disturbance and Landscape Patterns Affect Diversity Patterns of Aquatic Benthic Macroinvertebrates. *Journal of the North American Benthological Society*, 30, 284-295. <https://doi.org/10.1899/09-112.1>
- Manu, M. K., Ashiagbor, G., Seidu, I., Groen, T., Gyimah, T., & Toxopeus, B. (2023). Odonata as Bioindicator for Monitoring Anthropogenic Disturbance of Owabi Wetland Sanctuary, Ghana. *Aquatic Insects*, 44, 151-169. <https://doi.org/10.1080/01650424.2022.2108844>
- Mariottini, Y., De Wysiecki, M. L., Cepeda, R., Marinelli, C., Bardi, C. J., & Lange, C. E. (2024). Grasshopper (Orthoptera: Acridoidea) Diversity in the Pampas Region of Argentina: Status as Revealed by Long-Term Sampling. *Journal of Insect Conservation*, 28, 1265-1283. <https://doi.org/10.1007/s10841-024-00622-y>
- Marques, K. V. S., da Silva, P. G., & Frizzas, M. R. (2025). Seasonality-Related Variables Drive Taxonomic and Functional Diversity of Aquatic Beetle Larvae and Adults Differently in a Neotropical Savanna. *Biodiversity and Conservation*, 34, 2789-2807. <https://doi.org/10.1007/s10531-025-03096-9>
- Marta, S., Brunetti, M., Manenti, R., Provenzale, A., & Ficetola, G. F. (2021). Climate and Land-Use Changes Drive Biodiversity Turnover in Arthropod Assemblages over 150 Years. *Nature Ecology & Evolution*, 5, 1291-1300. <https://doi.org/10.1038/s41559-021-01513-0>
- Mensah, B. A., Kyerematen, R., Annang, T., & Adu-Acheampong, S. (2018). Influence of Human Activity on Diversity and Abundance of Insect in the Wetland Environment. *Bonorowo Wetlands*, 8, 33-41. <https://doi.org/10.13057/bonorowo/w080104>
- Menta, C., & Remelli, S. (2020). Soil Health and Arthropods: From Complex System to Worthwhile Investigation. *Insects*, 11, Article 54.

- <https://doi.org/10.3390/insects11010054>
- Miguel, T. B., Oliveira-Junior, J. M. B., Ligeiro, R., & Juen, L. (2017). Odonata (Insecta) as a Tool for the Biomonitoring of Environmental Quality. *Ecological Indicators*, *81*, 555-566. <https://doi.org/10.1016/j.ecolind.2017.06.010>
- Mondal, S., & Mondal, A. (2024). Diversity of Insect Fauna in Three Selected Sacred Groves of Murshidabad District, West Bengal, India. *Journal of Entomology and Zoology Studies*, *12*, 162-171. <https://doi.org/10.22271/j.ento.2024.v12.i3b.9330>
- Morilla, M., Serpa, K. V., Ogasawara, M. E. K., Piggott, J. J., Penk, M., Moretti, M. S. et al. (2026). Individual and Combined Effects of Shading and Habitat Complexity on Benthic Macroinvertebrate Communities in a Mesocosm Stream Experiment (São Paulo, Brazil). *Restoration Ecology*, e70376. <https://doi.org/10.1111/rec.70376>
- Mukherjee, S., Ghosh, L. K., & Debnath, N. (2018). A Preliminary Study on Insect Faunal Diversity in Sacred Groves of South Park Street Cemetery and St. Paul's Cathedral, Kolkata. *Journal of Experimental Zoology India*, *21*, 476-471.
- Mukweho, V. O., Pryke, J. S., & Roets, F. (2017). Habitat Preferences of the Invasive Harlequin Ladybeetle *Harmonia axyridis* (Coleoptera: Coccinellidae) in the Western Cape Province, South Africa. *African Entomology*, *25*, 86-97. <https://doi.org/10.4001/003.025.0086>
- Nganso, T. B., Kyerematen, R., & Obeng-Ofori, D. (2012). Review of Biodiversity in Sacred Groves in Ghana and Implications for Conservation. *Current Trends in Ecology*, *3*, 1-10.
- Nguyen, H. V., & Nguyen, H. T. T. (2025). Seasonal Variation of Aquatic Insect Biotic Indices and Their Relationship with Water Quality in the Tropical Mong Sen Stream, Lao Cai Province, Vietnam. *Egyptian Journal of Aquatic Biology and Fisheries*, *29*, 207-224. <https://doi.org/10.21608/ejabf.2025.429317.6709>
- Nicacio, G., & Juen, L. (2015). Chironomids as Indicators in Freshwater Ecosystems: An Assessment of the Literature. *Insect Conservation and Diversity*, *8*, 393-403. <https://doi.org/10.1111/icad.12123>
- Nnoli, H., Kyerematen, R., Adu-Acheampong, S., & Hynes, J. (2019). Change in Aquatic Insect Abundance: Evidence of Climate and Land-Use Change within the Pawmpawm River in Southern Ghana. *Cogent Environmental Science*, *5*, Article 1594511. <https://doi.org/10.1080/23311843.2019.1594511>
- Ntiama-Baidu, Y. (1995). *Indigenous versus Introduced Biodiversity Conservation Strategies: The Case of Protected Area Systems in Ghana*. Biodiversity Support Programme (Issues in African Biodiversity No. 1).
- Ohler, K., Schreiner, V. C., Link, M., Liess, M., & Schäfer, R. B. (2023). Land Use Changes Biomass and Temporal Patterns of Insect Cross-Ecosystem Flows. *Global Change Biology*, *29*, 81-96. <https://doi.org/10.1111/gcb.16462>
- Omokunle, A. B. (2024). Environmental Evaluation of a Tropical River Based on Aquatic Insect Communities and Water Quality Indicators. *Entomology Letters*, *4*, 8-16. <https://doi.org/10.51847/joblkojcre>
- Oppong, K. R. (2023). *Traditional Environmental Ethical Values on the Protection of Traditionally Protected Areas among the Asante Sekyere People*. Master's Thesis, University of Cape Coast.
- Osei, M. K., Aboagye, L. M., Ofori, P., & Annor, B. (2018). Biodiversity in Ghana. In T. Pullaiah (Ed.), *Global Biodiversity* (pp. 81-111). Apple Academic Press. <https://doi.org/10.1201/9780429469800-4>
- Pallottini, M., Goretti, E., Argenti, C., La Porta, G., Tositti, L., Dinelli, E. et al. (2023). But-

- terflies as Bioindicators of Metal Contamination. *Environmental Science and Pollution Research*, 30, 95606-95620. <https://doi.org/10.1007/s11356-023-28930-x>
- Parikh, G., Rawtani, D., & Khatri, N. (2021). Insects as an Indicator for Environmental Pollution. *Environmental Claims Journal*, 33, 161-181. <https://doi.org/10.1080/10406026.2020.1780698>
- Patole, S. N. (2022). Sacred Groves: Bridging Tradition and Biodiversity Conservation. *Vidyabharati International Interdisciplinary Research Journal*, 14, 1-5. <https://www.viirj.org/vol14issue1/16.pdf>
- Pedrosa Guimarães, L., Henrique Monteiro do Amaral, P., & da Gama Alves, R. (2025). Temporal Dynamics of Aquatic Insects and Oligochaetes in Tropical Springs. *Inland Waters*, 15, 1-11. <https://doi.org/10.1080/20442041.2025.2517112>
- Phillips, I., Jardine, T. D., Lindenschmidt, K., Westbrook, C., & Pomeroy, J. (2023). Nelson and Churchill River Basins. In M. D. Delong, T. D. Jardine et al. (Eds.), *Rivers of North America* (pp. 794-834). Elsevier. <https://doi.org/10.1016/b978-0-12-818847-7.00021-5>
- Ray, R., & Ray, A. (2020). Medicinal Practices of Sacred Natural Sites: A Socio-Religious Approach for Successful Implementation of Primary Healthcare Services. *Ethnobotany Research and Applications*, 20, 1-46. <https://doi.org/10.32859/era.20.34.1-46>
- Resh, V. H., Lévêque, C., & Statzner, B. (2004). Long-Term, Large-Scale Biomonitoring of Theunknown: Assessing the Effects of Insecticides to Control River Blindness (Onchocerciasis) in West Africa. *Annual Review of Entomology*, 49, 115-139. <https://doi.org/10.1146/annurev.ento.49.061802.123231>
- Rodríguez-Tapia, G., Prieto-Amparán, J. A., & Córdoba-Aguilar, A. (2022). Linking Potential Habitats of Odonata (Insecta) with Changes in Land Use/Land Cover in Mexico. *European Journal of Entomology*, 119, 272-284. <https://doi.org/10.14411/eje.2022.029>
- Samways, M. J. (2008). *Dragonflies and Damselflies of South Africa (No. 70)*. Pensoft Publishers.
- Sánchez-Argüello, R., Cornejo, A., Pearson, R. G., & Boyero, L. (2010). Spatial and Temporal Variation of Stream Communities in a Human-Affected Tropical Watershed. *Limnologia*, 40, 150-158.
- Sankone, C., Bedwell, C., & McCreadie, J. (2023). Regional B-Diversity of Stream Insects in Coastal Alabama Is Correlated with Stream Conditions, Not Distance among Sites. *Insects*, 14, Article 847. <https://doi.org/10.3390/insects14110847>
- Santana, H., Silva, L., Pereira, C., Simião-Ferreira, J., & Angelini, R. (2015). The Rainy Season Increases the Abundance and Richness of the Aquatic Insect Community in a Neotropical Reservoir. *Brazilian Journal of Biology*, 75, 144-151. <https://doi.org/10.1590/1519-6984.09213>
- Santos, L. R., & Rodrigues, M. E. (2022). Dragonflies (Odonata) in Cocoa Growing Areas in the Atlantic Forest: Taxonomic Diversity and Relationships with Environmental and Spatial Variables. *Diversity*, 14, Article 919. <https://doi.org/10.3390/d14110919>
- Scholtz, C., & Holm, E. (1985). *Insects of Southern Africa*. Butterworths Professional Publishers.
- Sharma, M., & Sharma, N. (2017). Suitability of Butterflies as Indicators of Ecosystem Condition: A Comparison of Butterfly Diversity across Four Habitats in Gir Wildlife Sanctuary. *International Journal of Advanced Research in Biological Sciences (IJARBS)*, 2, 43-53. <https://doi.org/10.22192/ijarbs.2017.04.03.005>
- Simaika, J. P., & Macadam, C. R. (2024). Insect Conservation in Streams and Rivers: Con-

- servation Threats and Solutions. In J. S. Pryke, M. J. Samways, T. R. New, P. Cardoso, & R. Gaigher (Eds.), *Routledge Handbook of Insect Conservation* (pp. 344-356). Routledge.
<https://doi.org/10.4324/9781003285793-31>
- Stanford, J. A., & Ward, J. V. (1983). Insect Species Diversity as a Function of Environmental Variability and Disturbance in Stream Systems. In J. R. Barnes, & G. W. Minshall (Eds.), *Stream Ecology* (pp. 265-278). Springer.
https://doi.org/10.1007/978-1-4613-3775-1_11
- Suhri, A. G. M., Jafar, J., Rahmah, M. H., Salatnaya, H., & Utami, R. B. (2025). Diversity and Ecological Role of Aquatic Insects as Bioindicators of Water Quality in Tropical Streams. *Egyptian Journal of Aquatic Biology and Fisheries*, *29*, 403-418.
<https://doi.org/10.21608/ejabf.2025.415515.6435>
- Susanto, M. A. D., Firdhausi, N. F., & Bahri, S. (2023). Diversity and Community Structure of Dragonflies (Odonata) in Various Types of Habitat at Lakarsantri District, Surabaya, Indonesia. *Journal of Tropical Biodiversity and Biotechnology*, *8*, Article 76690.
<https://doi.org/10.22146/jtbb.76690>
- Tan, C., Sheng, T., Wang, L., Mbaio, E., Gao, J., & Wang, B. (2021). Water-Level Fluctuations Affect the Alpha and Beta Diversity of Macroinvertebrates in Poyang Lake, China. *Fundamental and Applied Limnology*, *194*, 321-334.
<https://doi.org/10.1127/fal/2020/1297>
- Tatay, J., & Merino, A. (2023). What Is Sacred in Sacred Natural Sites? A Literature Review from a Conservation Lens. *Ecology and Society*, *28*, Article No. 12.
<https://doi.org/10.5751/es-13823-280112>
- Tubić, B., Andjus, S., Zorić, K., Vasiljević, B., Jovičić, K., Čanak Atlagić, J. et al. (2024). Aquatic Insects (Ephemeroptera, Plecoptera and Trichoptera) Metric as an Important Tool in Water Quality Assessment in Hilly and Mountain Streams. *Water*, *16*, Article 849.
<https://doi.org/10.3390/w16060849>
- Twining, C. W., Brenna, J. T., Lawrence, P., Winkler, D. W., Flecker, A. S., & Hairston, N. G. (2019). Aquatic and Terrestrial Resources Are Not Nutritionally Reciprocal for Consumers. *Functional Ecology*, *33*, 2042-2052. <https://doi.org/10.1111/1365-2435.13401>
- Vannote, R. L., Minshall, G. W., Cummins, K. W., Sedell, J. R., & Cushing, C. E. (1980). The River Continuum Concept. *Canadian Journal of Fisheries and Aquatic Sciences*, *37*, 130-137. <https://doi.org/10.1139/f80-017>
- Vero, S. E., & Snell, M. A. (2025). Role of Riparian Stream Shading on Stream Function and Implications for Catchment Management. *Biology and Environment: Proceedings of the Royal Irish Academy*, *125*, 29-41. <https://doi.org/10.1353/bae.2025.a955352>
- Wang, L., Li, J., Tan, L., & Han, B. (2023). Seasonal Patterns of Functional Alpha and Beta Redundancies of Macroinvertebrates in a Disturbed (Sub)Tropical River. *Ecological Indicators*, *146*, Article 109777. <https://doi.org/10.1016/j.ecolind.2022.109777>
- Wani, R. F. C., & Shah, I. M. (2024). Impact of Environmental Pollution on Diversity of Insects: A Case Study of Some Major Insect Orders. In Y. A. Hajam, S. H. Parey, & R. A. Bhat (Eds.), *Insect Diversity and Ecosystem Services* (pp. 17-53). Apple Academic Press.
<https://doi.org/10.1201/9781003471196-2>
- Windolowski, S. N. (2024). *The Thermal Response of Small Streams to Atmospheric Heat Waves in the Oregon Coast Range*. Master's Thesis.
- Worthen, W. B., & Guevara-Mora, M. (2024). The Effects of Light Environment on Adult Odonate Communities in Disturbed and Intact Forest: The Importance of Small-Scale Effects. *Diversity*, *16*, Article 557. <https://doi.org/10.3390/d16090557>
- Yang, Q., Wei, Z., Qiu, X., & Zhao, Z. (2025). Beta Diversity Patterns and Drivers of Macroinvertebrate Communities in Major Rivers of Ningxia, China. *Animals*, *15*, Article

2034. <https://doi.org/10.3390/ani15142034>

Yuen, E. Y. L., & Dudgeon, D. (2016). The Magnitude and Seasonality of Aquatic Insect Subsidies to Tropical Stream Riparia in Hong Kong. *Aquatic Sciences*, *78*, 655-667.

<https://doi.org/10.1007/s00027-015-0455-y>

Zhang, F., Tang, X., Lin, P., Shang, K., Han, Y., Liu, X. et al. (2025). Taxonomic and Functional Diversity of Macroinvertebrate Community along the Elevation Gradient and in Different Seasons in the Upper Jinsha River (Qinghai-Xizang Plateau). *Global Ecology and Conservation*, *62*, e03703. <https://doi.org/10.1016/j.gecco.2025.e03703>

Zhao, L., Gao, R., Liu, J., Liu, L., Li, R., Men, L. et al. (2023). Effects of Environmental Factors on the Spatial Distribution Pattern and Diversity of Insect Communities along Altitude Gradients in Guandi Mountain, China. *Insects*, *14*, Article 224.

<https://doi.org/10.3390/insects14030224>

Ziblila, M. H., Kaba, J. S., Yamoah, F. A., Acquaye, A. A., & Hashmiu, I. (2025). Sacred Groves for Enhanced Climate Mitigation: Towards a Framework for Managing Unintended Externalities of Environmental Systems. *Environmental Development*, *57*, Article 101351. <https://doi.org/10.1016/j.envdev.2025.101351>

Zina, V., Ordeix, M., Franco, J. C., Ferreira, M. T., & Fernandes, M. R. (2021). Ants as Bioindicators of Riparian Ecological Health in Catalonian Rivers. *Forests*, *12*, Article 625.

<https://doi.org/10.3390/f12050625>